

# Motif-Based Protein Sequence Classification Using Neural Networks

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## ABSTRACT

We present a system for multi-class protein classification based on neural networks. The basic issue concerning the construction of neural network systems for protein classification is the sequence encoding scheme that must be used in order to feed the neural network. To deal with this problem we propose a method that maps a protein sequence into a numerical feature space using the matching scores of the sequence to groups of conserved patterns (called motifs) into protein families. We consider two alternative ways for identifying the motifs to be used for feature generation and provide a comparative evaluation of the two schemes. We also evaluate the impact of the incorporation of background features (2-grams) on the performance of the neural system. Experimental results on real datasets indicate that the proposed method is highly efficient and is superior to other well-known methods for protein classification.

**Key words:** protein sequence classification, neural networks, probabilistic motifs, MEME algorithm, motif-based features.

## 1. INTRODUCTION

PROTEIN SEQUENCE CLASSIFICATION CONSTITUTES an important problem in biological sciences for annotating new protein sequences and detecting close evolutionary relationships among sequences. It deals with the assignment of sequences to known categories based on homology detection properties (sequence similarity). In several studies, protein classification has been examined at various levels, according to a top-down hierarchy in molecular taxonomy, consisting of superfamilies, families, and subfamilies (Dayhoff *et al.*, 1978). Throughout this paper, we will use the terms *family* (or *subfamily*) and *class* interchangeably to denote any collection of sequences that are presumed to share common characteristics and belong to the same category.

Various approaches have been developed for solving the protein classification problem. Most of them are based on appropriately modeling protein families, either directly or indirectly. Direct modeling techniques use a training set of sequences to build a model that characterizes the family of interest. Hidden Markov models (HMMs) are a widely used probabilistic modeling method for protein families (Durbin *et al.*, 1998) that provides a probabilistic measurement (score) of how well an unknown sequence fits to a family. Indirect techniques use direct models as a preprocessing tool in order to extract useful sequence features. In this way,

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sequences of variable length are transformed into fixed-length input vectors that are subsequently used for training discriminative models, such as neural networks.

In protein sequences, *motifs* or *patterns* enclose significant homologous attributes, since they correspond to conserved regions in protein families holding useful structural and functional biological properties. They can be considered as islands of amino acids conserved in the same order of a given family. Therefore, they can be seen as local features characterizing the sequences. Motifs can be either deterministic or probabilistic (Brázma *et al.*, 1998; Rigoutsos *et al.*, 2000). Deterministic motifs follow grammatical inference properties in order to syntactically describe conserved regions of homologous sequences. The PROSITE database (Hofmann *et al.*, 1999) represents a large collection of such motifs used to identify protein families. On the other hand, probabilistic motifs are more flexible models, and they provide a probabilistic matching score of a sequence to a motif. The BLOCKS database (Henikoff and Henikoff, 1994) is an example of ungapped probabilistic motifs. In any case, motif models are suitable for constructing efficient similarity score functions that can be subsequently used as local features for protein classification. An example is presented by Ma and Wang (2000), and by Wang *et al.* (2001) where motif-based local features are produced based on the minimum description length (MDL) principle for the case of deterministic motif models.

The *background* information also constitutes another source for extracting features from sequence data. The determination of the background features, also defined as *global* features, is usually made by using the *2-gram* encoding scheme that counts the occurrences of two consecutive amino acids in protein sequences (Wang *et al.*, 2001). In the case of protein sequences (generated from the alphabet of the 20 aminoacids), there are 400 possible 2-grams that produce a large feature space. A recent approach (Almeida and Vinga, 2002) proposes a scheme for globally encoding sequences, where each amino acid character is initially represented as a unique binary number with  $n$  bits ( $n = 5$  for the 20 aminoacids) and then each sequence is mapped into a position inside the  $n$ -dimensional hypercube.

In this paper, we focus on building efficient neural classifiers for discriminating multiple protein families by using appropriate local features that have been extracted by efficient probabilistic motif models. As motifs constitute family diagnostic signatures, our aim is to exploit this information by constructing a neural network scheme that exploits motif-based (local) features.

The proposed method can be considered as combining an unsupervised with a supervised learning technique. Starting by applying a motif-discovery (unsupervised) algorithm, we identify probabilistic motifs in a training set of multiclass sequences. This can be achieved in two alternative ways: applying the algorithm for motif discovery either to each family training set separately (*class-dependent* motifs), or to the whole dataset of training sequences (*class-independent* motifs). The discovered motifs are then used to convert each sequence to a numerical input vector that subsequently can be applied to a typical feed-forward neural network. Using a Bayesian regularization training technique, the neural network parameters are adjusted, and therefore a classifier is obtained suitable for predicting the family of an unlabeled sequence.

The next section provides a brief overview of statistical and neural techniques proposed for classifying biological sequences, while Section 3 describes the proposed method. Experimental results obtained using several sets of protein families are presented in Section 4, along with a comparison with other known protein classification approaches. Finally, Section 5 summarizes the proposed classification scheme and specifies directions for future research.

## 2. PROTEIN CLASSIFICATION METHODS

One class of methods for protein sequence classification work directly with sequence information and establish classification criteria based on sequence homology properties. In the general scheme, a representative set of sequences is selected for each protein family and used to build an appropriate model for each family. The classification of an unknown sequence is made by selecting the family that best matches according to the model homology mechanism. This can be considered as a simple *nearest neighbor* scheme that ranks sequence similarities and selects the best ranking.

The popular BLAST tool (Altschul *et al.*, 1990) represents the simplest nearest neighbor approach and exploits pairwise local alignments to measure sequence similarity. The BLAST technique compares protein

queries with a protein database of labeled sequences and produces normalized alignment scores for each comparison by calculating the corresponding expectation values ( $E$ -values). The classification procedure is based on the selection of the label of the sequence that produces the best pairwise alignment score (i.e., minimum  $E$ -value).

Another type of direct modeling methods is based on hidden Markov models (HMMs) (Durbin *et al.*, 1998; Karplus *et al.*, 1998). After constructing an HMM for each family, protein queries can be easily scored against all established HMMs by calculating the log-likelihood of each model for the unknown sequence and then selecting the class label of the most likely model.

The Motif Alignment and Search Tool (MAST) (Bailey and Gribskov, 1998) is based on the combination of multiple motif-based statistical score values. According to this scheme, groups of probabilistic motifs discovered by the MEME algorithm (Bailey and Elkan, 1994), are used to construct protein profiles for the families of interest. The MAST algorithm successively estimates the significance of the match of a query sequence to a family model as the product of the  $p$ -values of each motif match score. This measure (called  $E$ -value) can then be used to select the family of the unknown sequence.

Neural network schemes for protein classification consist of two stages: the *encoding* stage, where discriminative numerical features are computed for each training sequence, and the *decision* stage, where the created feature vectors are used as input vectors to a neural network classifier. Various encoding schemes have been proposed in the literature that produce numerical features in the encoding stage based on the calculation of background features (global sequence homology) and local features (locally conserved family information) embedded in motifs. In the decision stage, feed-forward neural networks have been used trained either through back-propagation (Wu *et al.*, 1996) or using Bayesian regularization (Ma and Wang, 2000; Wang *et al.*, 2001). These approaches are characterized by the enormous size of the extracted input vectors, the imbalance between global and local features (more emphasis on global features), and the need for large training sets (since the number of network inputs is very large). For example, in Ma and Wang (2000) and in Wang *et al.* (2001) only one feature was responsible for carrying local information, while all the others were produced by the 2-grams encoding scheme (background features).

Support vector machines (SVMs) (Vapnik, 1979) have been also applied to protein homology detection problems. Such an approach, which has been introduced by Logan *et al.* (2001), feeds probabilistic score values from all motifs available (nearly 10,000) in the BLOCKS database (Henikoff and Henikoff, 1994) into an SVM classifier. Obviously, this scheme uses only local features, but the dimensionality of the input space is extremely high. Another method has been proposed by Jaakkola *et al.* (2000) and by Karchin *et al.* (2002) that combines hidden Markov models (HMMs) and SVMs for detecting remote protein homologies. In particular, an HMM is first trained to model a protein family, and then the observed probabilities (in the log space) of each sequence with respect to each parameter of the HMM are calculated. The obtained gradient-log-probability vectors are applied to an SVM to identify the decision boundary between the family and the rest of the protein universe.

### 3. THE PROPOSED METHOD

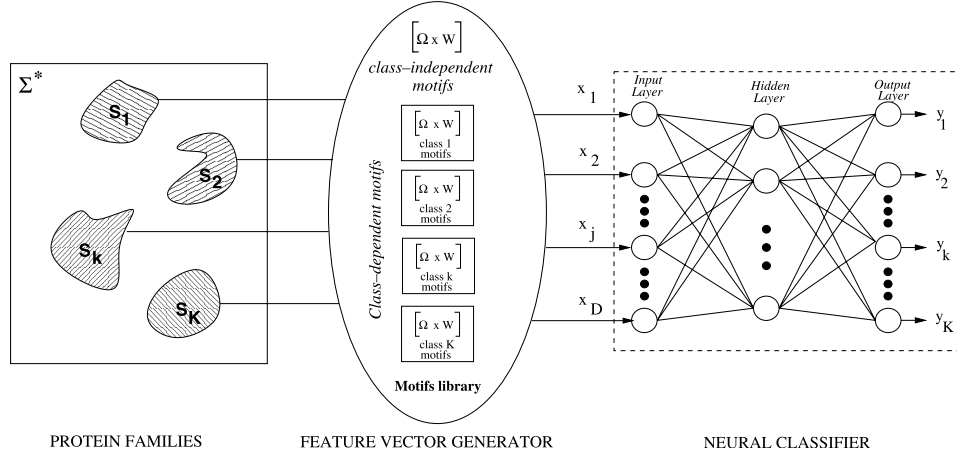
This paper studies the problem of classifying a set of  $N$  protein sequences  $\mathbf{S} = \{\mathcal{S}_i, i = 1, \dots, N\}$  into  $K$  classes. The set  $\mathbf{S}$  is a union of positive example datasets  $\mathcal{S}_i$  from  $K$  different classes, i.e.,  $\mathbf{S} = \{\mathcal{S}_1 \cup \dots \cup \mathcal{S}_K\}$ , and can be seen as a subset of the complete set of all possible sequences over the amino acid alphabet ( $\mathbf{S} \subseteq \Sigma^*$ ).

Figure 1 illustrates the architecture of the proposed protein classification scheme. It consists of a search tool (unsupervised learning) for discovering probabilistic motifs in a set of  $K$  protein families, a feature vector generator that converts protein sequences into feature vectors, and a decision module (neural network) for assigning a protein family to each input sequence. The following subsections describe in detail the major building blocks of the proposed architecture.

**F1**

#### 3.1. Using motifs for feature generation

Consider a finite alphabet consisting of set of characters  $\Sigma = \{\alpha_1, \dots, \alpha_\Omega\}$  ( $\Omega = 20$  for protein sequences). We can probabilistically model a contiguous (ungapped) motif  $M_j$  of length  $W_j$  using a



**FIG. 1.** The architecture of the proposed classification scheme.

position weight matrix ( $PWM_j$ ) that follows a multinomial character distribution. Each column ( $l$ ) of the matrix corresponds to a position  $l$  in the motif sequence ( $l = 1, \dots, W_j$ ), where the column elements provide the probability of each character of the alphabet  $p_{\alpha_{\xi},l}$  ( $\xi = 1, \dots, \Omega$ ) to appear in that position.

Let  $s_p = a_{p,1} \dots a_{p,W_j}$  denote a segment of a sequence  $S$  beginning at position  $p$  and ending at position  $p + W_j - 1$ . This represents a subsequence of length  $W_j$ . Totally, there are  $L - W_j + 1$  such subsequences for a sequence  $S$  of length  $L$ . Then, we can define the probability that  $s_p$  matches the motif  $M_j$ , or alternatively, has been generated by the model  $PWM_j$  corresponding to that motif, using the following equation:

$$P(s_p|M_j) = \prod_{l=1}^{W_j} p_{a_{p,l},l}. \quad (1)$$

A major advantage of using the probabilistic matrix  $PWM_j$  is the ability to compute the corresponding position-specific score matrix ( $PSSM_j$ ) in order to score a sequence. The  $PSSM_j$  is a log-odds matrix calculating the logarithmic ratio  $r_{\alpha_{\xi},l}$  of the probabilities  $p_{\alpha_{\xi},l}$  suggested by the  $PWM_j$  and the corresponding general relative frequencies of aminoacids  $\rho_{\alpha_{\xi}}$  in the family.<sup>1</sup> According to the definition of  $PSSM_j$ , the score value  $f_j(s_p)$  of a subsequence  $s_p$  of a sequence  $S$  can be defined as

$$f_j(s_p) = \sum_{l=1}^{W_j} \log \left( \frac{p_{a_{p,l},l}}{\rho_{a_{p,l}}} \right) = \sum_{l=1}^{W_j} r_{a_{p,l},l}. \quad (2)$$

At the sequence level, the score value of a protein sequence  $S$  against a motif  $M_j$  can be determined as the maximum value among all scores of the possible subsequences of  $S$ , i.e.,

$$f_j(S) = \max_{1 \leq p \leq L - W_j + 1} f_j(s_p). \quad (3)$$

It must be noted that it is possible to adopt other definitions for scoring a sequence, such as setting scores below a certain threshold equal to zero (Bailey and Gribskov, 1998).

If we assume that we have discovered a group of  $D$  motifs in the set of sequences  $\mathbf{S}$ , we can generate a  $D$ -dimensional numerical feature space and map each sequence  $S_i$  into a vector  $\mathbf{x}_i$  in the  $D$ -dimensional feature space by calculating the score values  $x_{ij} = f_j(S_i)$  ( $j = 1, \dots, D$ ) for each of the  $D$  motif models.

<sup>1</sup>The general relative frequencies of amino acids indicate the background information in a protein family and can be presented as a probabilistic vector  $\rho$  of size  $\Omega = 20$ .

### 3.2. Finding probabilistic motifs in protein sequences

Several approaches have been proposed for discovering probabilistic motifs in a set of unaligned biological sequences. CONSENSUS (Hertz and Stormo, 1999), the Gibbs sampler (Lawrence *et al.*, 1993), and MEME (Bailey and Elkan, 1994) are examples of such methods that identify multiple shared motifs in protein families. We have selected the MEME approach for the motif identification component of our strategy, since it has been widely used in biological applications and directly extracts position-specific score matrices. Below, we briefly describe this algorithm and propose two ways to integrate it in our classification system.

The MEME algorithm follows an iterative procedure, which applies at each iteration a two-component mixture model to discover one motif of length  $W$ . In the two-component model, one component describes the motif (ungapped common subsequences of length  $W$ ) while the other component models the background information. Multiple motifs can be found by sequentially fitting the two-component model to the set of sequences that remain after removing the sequences containing occurrences of the already identified motifs.

In particular, MEME (Bailey and Elkan, 1994) uses the Expectation Maximization (EM) algorithm (Dempster *et al.*, 1977) to maximize the log-likelihood function of the two-component mixture model, i.e., to estimate the elements of the corresponding position weight matrix.<sup>2</sup> Furthermore, MEME provides a strategy for locating efficient initial parameter values in order to prevent the EM algorithm from getting stuck in local optima (Bailey and Elkan, 1994). The  $D$  motif models  $PWM_j$  ( $j = 1, \dots, D$ ) discovered by MEME can be of either fixed or variable length  $W_j$ . In our experimental studies, both types of motifs will be examined to evaluate the impact of this decision on the performance of the neural classifier.

In order to discover a group of motifs from a multiclass training set of sequences (containing sequences of  $K$  classes), two alternative approaches can be followed. The first approach is to apply the MEME algorithm  $K$  times, *separately* to the training sequences of each protein family. Then, putting all the discovered  $K$  family profiles together, we can form the final group of  $D$  motifs. An alternative approach is to apply the motif-discovery algorithm only once to the total training set  $\mathbf{S}$ , ignoring class labels. In this way, we do not allow the algorithm to directly create  $K$  protein family profiles, but rather to discover  $D$  *class-independent* motifs.

The advantage of the second approach is the ability of taking into account local similarity measurements in the whole training set, without restricting the search procedure to a single class. Therefore, possible partial homologies among sequences from different families can be defined that may prove helpful for the classification task. On the other hand, a disadvantage of the class-independent approach is that the  $D$  discovered motifs may not be equally distributed among the  $K$  families. This may result in insufficient modeling of some families, thus leading to performance deterioration. During experiments, both motif-discovery strategies will be considered and evaluated.

### 3.3. Construction of a neural classifier

After discovering  $D$  motifs and constructing the  $D$ -dimensional feature space, the last stage in our methodology is to implement and train a feed-forward neural network that will be able to map the input vectors into the protein classes of interest. A typical network architecture is illustrated in Fig. 1. To construct the neural classifier, we use the training set  $\mathbf{X} = \{\mathbf{x}_i, \mathbf{t}_i\}$ ,  $i = 1, \dots, N$  consisting of positive examples  $\mathbf{x}_i$  from the set of  $K$  protein families. The target vector  $\mathbf{t}_i$  is a binary vector of size  $K$  indicating the class label of input  $\mathbf{x}_i$ ; i.e.,  $t_{ik} = 1$  if  $\mathbf{x}_i$  corresponds to a sequence  $S_i$  belonging to class  $k$ , and 0 otherwise. The output of the classifier is represented by the  $K$ -dimensional vector  $\mathbf{y}_i$  where component  $y_{ik}$  corresponds to class  $k$ . Based on this scheme, the predicted class  $h(\mathbf{x}_i)$  of an unlabeled feature vector  $\mathbf{x}_i$  corresponding to a query sequence  $S_i$  is given by the index of the output node with the largest value  $y_{ic}$ ; i.e.,

$$h(\mathbf{x}_i) = c : y_{ic} = \max_{1 \leq k \leq K} y_{ik} . \quad (4)$$

<sup>2</sup>The model used in our experiments assumes that there are zero or more nonoverlapping occurrences of the motif in each sequence of the dataset. Alternative models that can be used are the exactly one-occurrence-per-sequence and the zero-or-one-occurrence-per-sequence models.

Setting a threshold value  $\theta$  ( $\in [0, 1]$ ), we can restrict the classifiers' decision to only those input vectors whose maximum output value surpasses this threshold. In this case, we can write

$$h(\mathbf{x}_i, \theta) = c : y_{ic} = \max_{1 \leq k \leq K} y_{ik} \wedge y_{ic} \geq \theta . \quad (5)$$

Parameter  $\theta$  can be used to specify the sensitivity of the classifier.

In order to train the neural network, we used the Gauss–Newton Bayesian Regularization (GNBR) learning algorithm (Foresse and Hagan, 1997). This algorithm applies Bayesian regularization and implements a Gauss–Newton approximation to the Hessian matrix of the objective function.

In the Bayesian regularization framework, the objective function is formulated as the weighted sum of two terms: the sum of the squared errors ( $E_X$ ) and the sum of squares of the network weights ( $E_W$ ). Using Bayes' rule, the posterior probability distribution for the weights  $\mathbf{w}$  of the network given a training set  $\mathbf{X}$  can be written as follows:

$$P(\mathbf{w}|\mathbf{X}) = \frac{P(\mathbf{X}|\mathbf{w})P(\mathbf{w})}{P(\mathbf{X})} . \quad (6)$$

By properly choosing the prior distribution  $P(\mathbf{w})$  and the likelihood function  $P(\mathbf{X}|\mathbf{w})$ , we can obtain the following expression (Bishop, 1995; Foresse and Hagan, 1997) for the posterior distribution:

$$P(\mathbf{w}|\mathbf{X}) = \frac{1}{Z_F} \exp(-\beta E_X - \alpha E_W) = \frac{1}{Z_F} \exp(-F(\mathbf{w})), \quad (7)$$

where the  $Z_F$  corresponds to the normalizing factor that is independent of the weights.

Maximizing the above posterior distribution is equivalent to minimizing the regularized objective function  $F(\mathbf{w})$ :

$$F(\mathbf{w}) = \frac{\beta}{2} \sum_{i=1}^{N_X} \{\mathbf{y}_i - \mathbf{t}_i\}^2 + \frac{\alpha}{2} \sum_{j=1}^{N_W} w_j^2 , \quad (8)$$

where  $N_X$  and  $N_W$  represent the number of input vectors and network parameters, respectively. In order to estimate the normalizing factor  $Z_F$ , a Gaussian approximation can be used for the posterior distribution (MacKay, 1992) as obtained by the Taylor expansion of function  $F(\mathbf{w})$  around the minimum value of the posterior,  $\mathbf{w}_{MP}$ . This gives the following estimation (Bishop, 1995):

$$Z_F^*(\alpha, \beta) = \exp(-F(\mathbf{w}_{MP}))(2\pi)^{N_W/2} |\mathbf{H}|^{-1/2} , \quad (9)$$

where  $\mathbf{H}$  corresponds to the Hessian matrix of the regularized objective function and, therefore, optimal values for parameters  $\alpha$  and  $\beta$  at the minimum point  $\mathbf{w}_{MP}$  can be computed as follows:

$$\hat{\alpha} = \frac{\gamma}{2E_W(\mathbf{w}_{MP})} \quad \text{and} \quad \hat{\beta} = \frac{\gamma N_X}{2E_X(\mathbf{w}_{MP})} . \quad (10)$$

The quantity  $\gamma$  represents the effective number of network parameters  $\mathbf{w}$  and can be defined using the eigenvalues of  $H^{-1}$  as  $\gamma = N_W - 2\alpha \text{Tr} \mathbf{H}^{-1}$ . In cases where the number of effective parameters is equal to the actual ones ( $\gamma \approx N_W$ ), more hidden units must be added to the network. Furthermore, the GNBR algorithm follows a Gauss–Newton approximation method (Foresse and Hagan, 1997) for calculating the Hessian matrix of  $F(\mathbf{w})$  at the minimum point  $\mathbf{w}_{MP}$ , using the Levenberg–Marquardt optimization algorithm (Bishop, 1995). It must be noted that in our experiments, the best results for the GNBR algorithm were obtained by scaling the network inputs in the range  $[-1, 1]$ .

#### 4. EXPERIMENTAL RESULTS

Several experiments were conducted to evaluate the proposed method. The classification accuracy was measured by counting the sensitivity and specificity rates. In all  $K$ -class classification problems, each

TABLE 1. THE TWO PROSITE FAMILIES USED IN THE EXPERIMENTAL STUDY

Problem: PROSITE 1 ( $K = 6$ )			Problem: PROSITE 2 ( $K = 7$ )		
PROSITE family	Positive data	Training set (avg length of seqs)	PROSITE family	Positive data	Training set (avg length of seqs)
PS00030	302	20 (370)	PS00070	129	15 (558)
PS00038	289	20 (359)	PS00077	155	15 (502)
PS00061	317	20 (299)	PS00118	168	15 (127)
PS00198	300	20 (284)	PS00180	123	15 (408)
PS00211	574	30 (478)	PS00215	123	15 (321)
PS00301	386	20 (517)	PS00217	148	15 (490)
			PS00338	173	15 (212)

protein family  $\mathcal{S}_{||}$  ( $k = 1, \dots, K$ ) was randomly partitioned into training and test sequences, with the training set being only a small percentage (5–10%) of the family dataset. Using the training datasets, experiments have been carried out using the MEME algorithm to discover groups of motifs. Two cases were considered: in the first case, the MEME algorithm has been applied separately to each training set providing a group of  $D_k = 5$  class-dependent motifs for each family  $\mathcal{S}_{||}$ .<sup>3</sup> In the second case, the MEME algorithm was applied only once to the total training dataset (ignoring the class labels) to provide a group of  $D = 5 \times K$  class-independent motifs.

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In any case, the obtained final group of  $D$  motifs were used to transform each sequence of the dataset into a dataset with numerical  $D$ -dimensional feature vectors, denoted  $\mathbf{X}_s$  for the class-dependent case and  $\mathbf{X}_g$  for the class-independent case. Furthermore, we also experimented with the effect of the length  $W$  of the discovered motifs to the performance of the proposed classifier, by applying the MEME algorithm with either fixed or variable motif length. We selected  $W = 20$  for the first case and the range  $[10, 30]$  for the second case. In summary, we have considered four distinct cases considering the application of MEME: discovering either class-dependent or class-independent motifs with either fixed or variable motif length. Therefore, for each classification problem, four distinct neural classifiers will be constructed and tested.

To evaluate classification performance, ROC (receiver operating characteristic) analysis was used. More specifically, we used the ROC<sub>50</sub> curve which is a plot of the sensitivity as a function of false positives for various decision threshold values until 50 false positives are found.

For our experimental study, three real datasets were selected. In particular we have used protein families from the PROSITE database (Hofmann *et al.*, 1999), which is a large collection of protein families together with their characteristic (deterministic) motifs. Two datasets with  $K = 6$  (PROSITE 1) and  $K = 7$  (PROSITE 2) classes from the PROSITE database (Hofmann *et al.*, 1999) were selected, summarized in Table 1. Moreover, experiments have also been conducted on a dataset of G-protein coupled receptors (GPCR) (Horn *et al.*, 1998), that is, a superfamily of cell membrane proteins. The GPCR database is hierarchically classified into five major classes and their subfamilies (Horn *et al.*, 1998). We studied the problem of classifying subfamilies within the class A, since it dominates the whole GPCR database. As indicated by Karchin *et al.* (2002), the difficulty of recognizing GPCR subfamilies arises from the fact that the classification of the subfamilies has been made based on chemical properties rather than sequence homology. Therefore, members from different subfamilies may share strong homology, thus making their discrimination hard. Among 15 subfamilies consisting of class A, seven of them have been selected in our experimental study described in Table 2. The remaining eight subfamilies are of very small size, and it is difficult to construct an effective system for their discrimination. Details of the three datasets (family/subfamily names and their protein ID's) used in our experiments are given in the appendix.

T1

T2

#### 4.1. Local versus global features

In this series of experiments, we assessed the impact of using 2-grams (background features) on the performance of the proposed classification scheme. For a sequence  $S_i$  with length  $L_i$ , we define the feature

<sup>3</sup>Experiments with a greater number of motifs did not yield better classification performance.

TABLE 2. SEVEN FAMILIES FROM THE GPCR CLASS A USED IN THE EXPERIMENTAL STUDY

<i>Problem: GPCR (K = 7)</i>		
<i>GPCR Class A subfamily</i>	<i>Positive data</i>	<i>Training set (avg length of seqs)</i>
Amine	306	20 (485)
Peptide	654	30 (383)
Hormone	43	10 (378)
Rhodopsin	270	20 (358)
Olfactory	325	20 (317)
Prostanoid	43	10 (721)
Nucleotide-like	58	10 (348)

value  $g_{iq}$  for each 2-gram  $q$  with respect to this sequence as

$$g_{iq} = \frac{\mathcal{N}(q|S_i)}{L_i - 1}, \quad (11)$$

where  $\mathcal{N}(q|S_i)$  denotes the number of occurrence of the 2-gram feature  $q$  in the sequence  $S_i$ . Obviously, the above equation gives the relative frequency of a 2-gram feature in a sequence. In a training set  $\mathbf{S} = \{S_1, S_2, \dots, S_N\}$  of  $N$  sequences, we can ignore *redundant* 2-grams and consider only the  $N_g$  features  $g_{iq}$  that correspond to the most frequently occurring 2-grams. We select the  $N_g$  2-grams occurring in at least half of the training sequences and by computing the corresponding  $g_{iq}$  ( $q = 1, \dots, N_g$ ) values for each sequence  $S_i$ , we construct the corresponding feature vectors to be fed in the neural classifier.

Table 3 presents the dimensionality of the feature spaces obtained using 2-grams and motifs for each dataset used in the experiments. It must be noted that we can further reduce the dimensionality of the 2-gram feature vectors using standard dimension reduction techniques, such as principal component analysis (PCA).

To assess the impact of the several feature types on the performance of the classification system, we have considered five different datasets:

- $\mathbf{X}_S$ :  $D$  motif-based features separately identified for each family (class-dependent),
- $\mathbf{X}_G$ :  $D$  motif-based class-independent features,
- $\mathbf{X}_S \cup \mathbf{G}$ :  $D$  motif-based class-dependent features along with  $N_g$  2-gram features,
- $\mathbf{X}_G \cup \mathbf{G}$ :  $D$  motif-based class-independent features, along with  $N_g$  2-gram features
- $\mathbf{G}$ :  $N_g$  2-gram features.

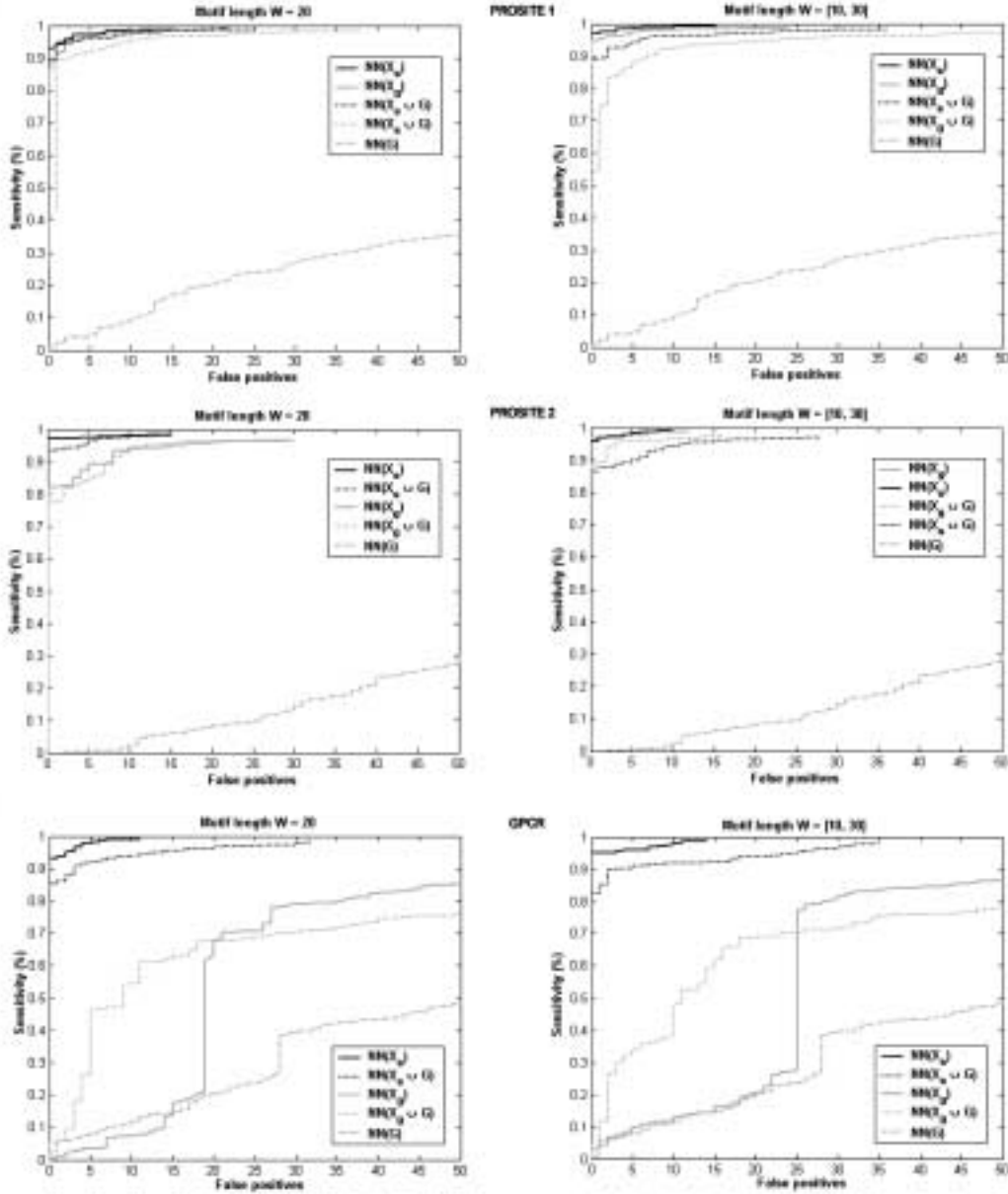
The neural network architecture had one hidden layer of either 10 (for the cases  $\mathbf{X}_S$  and  $\mathbf{X}_G$ ) or 20 nodes (for the other three cases).

Figure 2 displays the ROC<sub>50</sub> curves obtained after training the five neural classifiers in each of the three classification problems, respectively. For each problem, two different graphs are presented concerning

TABLE 3. THE NUMBER OF THE EXTRACTED MOTIF-BASED ( $D$ ) AND 2-GRAM ( $N_g$ ) FEATURES THAT CORRESPONDS TO EACH DATASET

<i>Problem</i>	<i><math>N_g</math> 2-gram features</i>	<i><math>D</math> motif-based features</i>
PROSITE 1	174	$5 \times 6 = 30$
PROSITE 2	285	$5 \times 7 = 35$
GPCR	152	$5 \times 7 = 35$

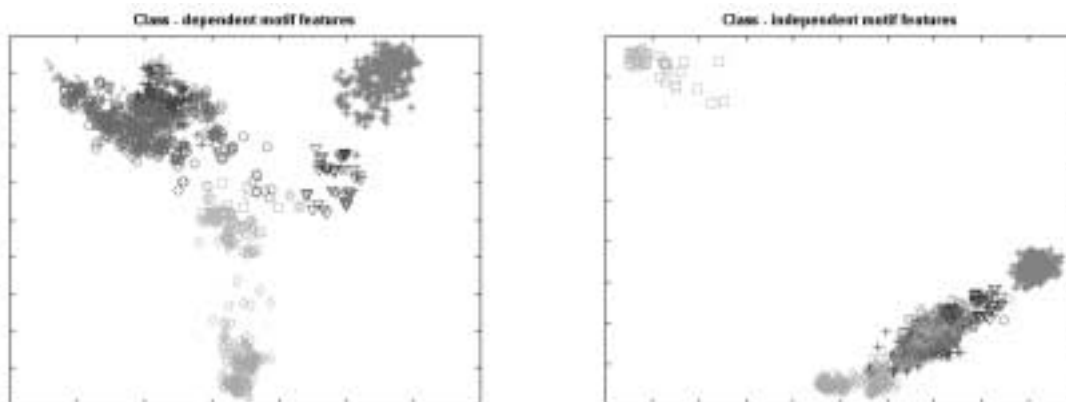




**FIG. 2.** ROC<sub>50</sub> curves illustrating the performance of the neural classifier on the three datasets using the five different feature vectors.

motifs of fixed length ( $W = 20$ ) and of variable length  $W \in [10, 30]$ . Obviously, motif-based features themselves constitute an excellent source of information able to generate significant features and lead to the construction of efficient classifiers. In all cases, the neural networks trained by mixed features (e.g.,  $NN(X_s \cup G)$ ) exhibit lower classification accuracy compared to the corresponding classifier trained with only motif-based features (e.g.,  $NN(X_s)$ ). Furthermore, the 2-grams features alone (case  $NN(G)$ ) do not seem to contain significant discriminant information.

Another observation that can be made from the ROC<sub>50</sub> curves in Fig. 2 is related to the performance of the neural classifier with class-dependent motifs (network  $NN(X_s)$ ) compared to that obtained with class-independent motifs (network  $NN(X_g)$ ). In almost all cases, we obtained better classification results with the network  $NN(X_s)$ . One explanation for this behavior is that, when searching for a specific number  $D$  of motifs in the whole training set (ignoring class labels), the algorithm may focus on some of the families



**FIG. 3.** The seven class regions in the GPCR dataset in the case of class-dependent and class-independent features. The data have been projected in two dimensions using PCA.

and leave the other families explored only partially. This possibly affects the satisfactory modeling of some families, since the discovered class-independent motifs may not be sufficient for describing them (only a few individual motifs are dedicated to this family). Experiments in the  $\mathbf{X}_g$  datasets with MEME have shown that the allocation of motifs in most cases was not equal for all the  $K$  families.

An example is shown in Fig. 3 that illustrates the constructed feature space of the  $\mathbf{X}_s$  and  $\mathbf{X}_g$  datasets in the case of the GPCR problem (seven classes), after projecting the 35-dimensional numerical to a two-dimensional space using PCA. It can be observed that in the case of class-dependent motifs the protein classes exhibit less overlap while in the reduced feature space of class-independent motifs there is a significant overlapping among class regions, thus making the discrimination harder. A selection of higher values of  $D$  probably would lead to better results for the class-independent case, but would simultaneously result in larger feature spaces or to the overestimation of some families.

**F3**

#### 4.2. Comparison with other approaches

We have also compared the neural classifier (with class-dependent motif-based features) with two other protein classification methods, namely, the MAST homology detection algorithm (Bailey and Gribskov, 1998) and the profile HMMs built using SAM, (Hughey and Krogh, 1996). In both MAST and SAM, each protein family (or subfamily) is transformed (indirectly or directly) into a probabilistic model-profile, and the test sequences are classified using the class of the profile with the best score value.

More specifically, the MAST procedure (Bailey and Gribskov, 1998) initially uses the MEME algorithm to discover groups of motifs separately for each one of the  $K$  protein families. For each sequence in the testing set, the MAST algorithm combines the calculated  $p$ -values and estimates the significance of the observed match (called  $E$ -value) of the sequence to each of the  $K$  groups of motifs.<sup>4</sup> Then the query sequence is assigned to the class with the minimum  $E$ -value. The SAM method (Hughey and Krogh, 1996) works in a similar way by building an HMM for each one of the  $K$  protein families (or subfamilies) instead of discovering groups of motifs.<sup>5</sup>

**FN4**

**FN5**

**F4**

Figure 4 provides comparative results from the application of the proposed neural classifier, MAST and SAM, to the three datasets. We have created five ROC curves for each method (number of false positives versus sensitivity for several threshold values) until 25 false positives were found ( $\text{ROC}_{25}$ ). The performance of the neural classifier and MAST was given by two curves respectively<sup>6</sup> concerning motifs of fixed ( $W = 20$ ) and variable length ( $W = [10, 30]$ ), while the last one corresponds to SAM performance.

**FN6**

<sup>4</sup>We use the *meme* and *mast* commands from the available MEME package v.3.0.4.

<sup>5</sup>We use the *buildmodel* and *hmmscore* commands from the available SAM package v.3.3.1.

<sup>6</sup>The curves for the neural classifier performance were the best plots from the corresponding  $\text{ROC}_{50}$  diagrams in Fig. 2.

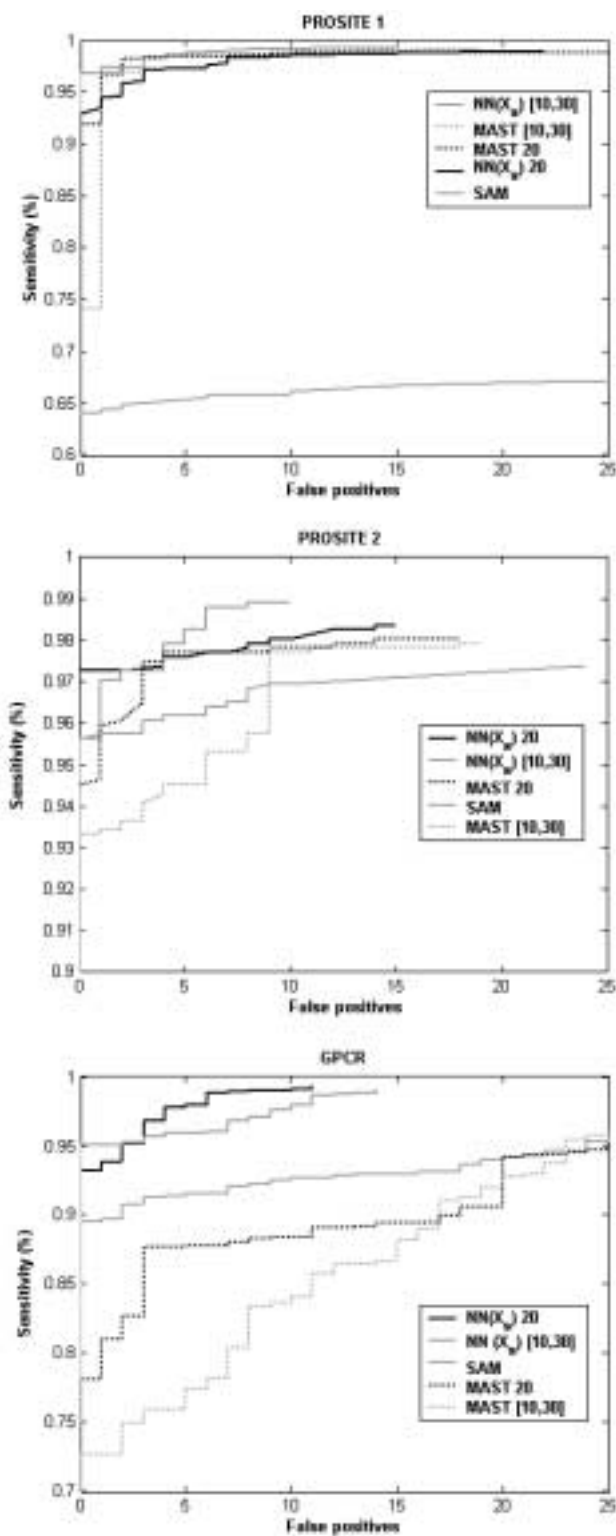


FIG. 4. ROC<sub>25</sub> curves for the three methods (neural (NN), MAST, and SAM) on the three datasets.

In the case of MAST and SAM methods, ROC curves were obtained by setting several  $E$ -value thresholds. When the lowest estimated  $E$ -value for a query sequence was greater than the threshold, then the test sequence was considered unclassified.

The superior classification of the proposed neural approach is obvious from the plotted curves in all problems, offering greater sensitivity rates with perfect specificity (zero false positives). For the GPCR dataset, which is more difficult to discriminate, the classification improvement is more clear: a sensitivity rate of 99.30% was measured with only 11 false positives, while the corresponding results for MAST and SAM are (95.76%, 25) and (95.38%, 25), respectively. It is also important to stress the higher accuracy that the neural scheme achieves compared with MAST (dot lines). Although these two methods use the same groups of motifs, our method seems to offer a more efficient scheme for combining the motif match scores compared to the combination of their  $p$ -values as suggested by MAST. In addition, the neural classifier achieves fewer false positives with higher sensitivity rates in all datasets concerning either fixed or variable motif length. Again, the improvement is more clear in the plots corresponding to the GPCR dataset.

Regarding more carefully the three selected datasets, they can be considered as three different types of protein sequence classification problems. In particular, the PROSITE 1 dataset consists of *diverse protein families* in the sense that their corresponding PROSITE motifs are not very specific (such as in the case of PS00030 and PS00198) and they can be found in sequences from a large number of protein families. Hence, this application can be seen as a diverse protein family recognition problem. On the other hand, the PROSITE 2 dataset consists of protein families with more specific PROSITE motifs that can be distinguished more easily. Finally, the third dataset, GPCR, is related to the recognition of protein subfamilies within a broader protein family domain sharing strong homology.

In all the above three types of protein sequences classification problems, our approach has shown a superior classification performance providing better results in comparison with the two other approaches. As illustrated in Fig. 4, the SAM method seems to be unsuccessful in recognizing diverse protein families (PROSITE 1 case), and the obtained classification rate was low (the individual classification error for each diverse family was about 50%). On the other hand, the performance of the MAST method was lower in the case of the GPCR subfamily recognition problem where sequences from different subfamilies share strong homology. Finally, in the case of recognizing simple protein families (PROSITE 2 dataset), all the three approaches provide similar classification rates, with the proposed neural scheme offering slightly better results.

## 5. CONCLUSIONS

In this paper, we have presented a neural network approach for the classification of protein sequences. The proposed methodology is motivated by the principle that in biological sequence analysis motifs can provide major diagnostic features for determining the class label of the unknown sequences. The method is implemented in two steps, where a preprocessing step (based on the MEME algorithm) is initially applied for discovering a group of probabilistic motifs appearing in the sequences. We have suggested and evaluated two alternative ways for motif discovery in a set of  $K$ -class sequences depending on whether the class labels are taken into account. Using the discovered motifs, a numerical feature vector is generated for each sequence by computing the matching score of the sequence to each motif. At the second stage of the proposed method, the extracted feature vectors are used as inputs to a feed-forward neural network trained using the Gauss–Newton Bayesian Regularization algorithm that provides the class label of a sequence.

Experiments were conducted on real datasets (using very small training sets), and comparisons were made with the MAST and SAM probabilistic methods. ROC curves were used as a performance indicator, and the experimental results clearly illustrate the superiority of the proposed neural system. In addition we have shown that background features do not constitute a useful source of information for the classification task since they do not lead to performance improvement.

In future work, more extensive experiments could be conducted to assess the performance of the method on specific protein superfamilies of important biological functions, as was the case with the GPCR dataset. Also, alternative methods could be implemented and tested, both in the classification stage (mixture models, SVMs, etc.) and in the motif discovery stage.

## APPENDIX: DATASETS

In the next tables proteins with bold ID's correspond to the training examples and the rest of them to the test set.

T4-T6

TABLE 4. DESCRIPTION OF THE PROSITE 1 DATASET

Family	Protein ID's
PS00030	<p><b>CB20-HUMAN GAR2-SCHPO HRB1-YEAST HS49-YEAST IF34-MOUSE NAB4-YEAST PAB3-ARATH RB27-DROME RN15-YEAST ROA1-MOUSE ROC3-NICSY RU17-HUMAN RU17-YEAST RU1A-DROME RU2B-HUMAN SFPQ-HUMAN U2AF-CAEEL U2AF-HUMAN U2AG-HUMAN K682-HUMAN A2BP-HUMAN A2BP-MOUSE ARP2-PLAFA ROAA-MOUSE CAZ-DROME CB20-XENLA CG79-HUMAN PM14-MOUSE CIRP-HUMAN CIRP-MOUSE CIRP-XENLA CPO-DROME CST2-HUMAN CSX1-SCHPO CTF1-SCHPO CUG1-HUMAN CUG1-MOUSE CWF5-SCHPO CYPE-DROME CYPE-HUMAN CYPE-MOUSE D111-ARATH ELAV-DROME ELAV-DROVI ELV1-HUMAN ELV1-MOUSE ELV2-HUMAN ELV2-MOUSE ELV3-HUMAN ELV3-MOUSE ELV4-HUMAN ELV4-RAT ELV4-RAT EWS-HUMAN EWS-MOUSE FCA-ARATH FUS-BOVIN FUS-HUMAN FUS-MOUSE G3B2-HUMAN G3B2-MOUSE G3BP-HUMAN G3BP-MOUSE G3BP-SCHPO GBP2-YEAST GR10-BRANA GRF1-HUMAN GRP1-SINAL GRP1-SORBI GRP2-SINAL GRP2-SORBI GRP7-ARATH GRP8-ARATH GRPA-MAIZE GRP-DAUCA IF34-CAEEL IF34-HUMAN IF34-SCHPO IF34-YEAST IF39-HUMAN IF39-SCHPO IF39-TOBAC IF39-YEAST IF4B-HUMAN IF4B-YEAST IF4H-HUMAN IF4H-MOUSE JSN1-YEAST LAA-XENLA LAB-XENLA LAH1-SCHPO LAH1-YEAST LA-AEDAL LA-BOVIN LA-DROME LA-HUMAN LA-MOUSE LA-RABIT LA-RAT MAT3-HUMAN MAT3-RAT MEI2-SCHPO MLO3-SCHPO MODU-DROME MSSP-HUMAN NAB3-YEAST NAM8-YEAST NGR1-YEAST NONA-DROME NOP3-YEAST NOP4-YEAST NOP8-YEAST NQ2-YEAST NR54-HUMAN NRD1-SCHPO NRD1-YEAST NRP1-YEAST NSR1-DROME NUCL-CHICK NUCL-HUMAN NUCL-MESAU NUCL-MOUSE NUCL-RAT NUCL-XENLA PAB1-HUMAN PAB1-MOUSE PAB2-ARATH PAB2-HUMAN PAB4-HUMAN PABS-ARATH PABP-DROME PABP-SCHPO PAB1-XENLA PABP-YEAST PABX-ARATH PES4-YEAST PRT1-PICAN PTB-HUMAN PTB-MOUSE PTB-PIG PTB-RAT PUB1-YEAST RB56-HUMAN RB87-DROME RB97-DROME RBM3-HUMAN RBM3-MOUSE RBM5-HUMAN RBM6-HUMAN RBM7-HUMAN RB8A-HUMAN RBM9-HUMAN RBMA-HUMAN RBMA-RAT RBMB-HUMAN RBMS-CHICK RBMS-HUMAN RBMS-MOUSE RBMS-XENLA RBP1-DROME RBP1-MOUSE RBP1-RAT RBP1-SYN3 RDP-HUMAN RDP-MOUSE RN24-SCHPO RNP1-YEAST RO21-XENLA RO22-XENLA RO31-XENLA RO32-XENLA ROA0-HUMAN ROA1-BOVIN ROA1-DROME ROA1-HUMAN ROA1-MACMU ROA1-RAT ROA1-SCHAM ROA1-XENLA ROA2-HUMAN ROA2-MOUSE ROA3-HUMAN ROAB-ARTSA ROC1-ARATH ROC1-NICPL ROC1-NICSY ROC1-SPIOL ROC2-ARATH ROC2-NICPL ROC2-NICSY ROC3-ARATH ROC4-NICSY ROC5-NICSY ROC-HUMAN ROC-RAT ROC-XENLA ROD-HUMAN ROD-RAT ROF-HUMAN ROG-HUMAN ROG-MOUSE ROH1-HUMAN ROH2-HUMAN ROL-HUMAN ROM-HUMAN ROR-HUMAN ROU2-HUMAN RS31-ARATH RS40-ARATH RS41-ARATH RT19-ARATH RU17-ARATH RU17-DROME RU17-MOUSE RU17-XENLA RU1A-HUMAN RU1A-XENLA RU1A-YEAST RX21-DROME S3B4-HUMAN SCE3-SCHPO SFR1-ARATH SFR1-HUMAN SFR2-CAEEL SFR2-CHICK SFR2-HUMAN SFR2-MOUSE SFR3-HUMAN SFR4-HUMAN SFR5-HUMAN SFR5-MOUSE SFR5-RAT SFR6-HUMAN SFR6-RABIT SFR7-HUMAN SFR9-HUMAN SFRB-HUMAN SFRB-MOUSE SQD-DROME SR55-DROME SRA4-HUMAN SRA4-RAT SRP1-SCHPO SSB1-YEAST SXL-CERCA SXL-CHRRU SXL-DROME SXL-DROSU SXL-MEGSC SXL-MUSDO TIA1-HUMAN TIA1-MOUSE TIAR-HUMAN TIAR-MOUSE TR2A-HUMAN TR2A-DROME TR2A-DROVI U2AF-CAEER U2AF-DROME U2AF-MOUSE U2AF-SCHPO U2AG-DROME U2AG-MOUSE U2AG-SCHPO U2R1-HUMAN U2R1-MOUSE U2R2-HUMAN U2R2-MOUSE WHI3-YEAST XMS2-DROME Y117-HUMAN YAC4-SCHPO YAG3-SCHPO YAS9-SCHPO YBFI-YEAST YBLC-SCHPO YCQ9-SCHPO YD3D-SCHPO YDB2-SCHPO YDC1-SCHPO CWF2-SCHPO YDR6-SCHPO YFK2-YEAST YG5B-YEAST YH4C-YEAST YHHS-YEAST YI51-YEAST YI55-YEAST YI59-YEAST YKVA-YEAST YLFI-CAEEL YML7-YEAST YN26-YEAST YN8T-YEAST YNL0-YEAST YNR5-YEAST YP68-YEAST YP85-CAEEL YQ01-CAEEL YQ04-CAEEL YQ0A-CAEEL YQ0C-CAEEL YRA1-YEAST YS07-CAEEL YSX2-CAEEL</b></p>
PS00038	<p><b>AHR-RAT ARRS-MAIZE CBF1-YEAST DA-DROME ESM7-DROME HEN2-MOUSE HES3-MOUSE MAD4-MOUSE MITF-RAT MXII-BRARE MYC-HYLLA MYF6-HUMAN MYOD-BRARE NDF1-MESAU SIM1-MOUSE TAL2-MOUSE TAL-HUMAN TF21-MOUSE TWS1-HUMAN USF2-RAT AHR-HUMAN AHR-MOUSE ARLC-MAIZE ARN2-HUMAN ARN2-MOUSE ARNT-DROME ARNT-HUMAN ARNT-MOUSE ARNT-RABIT ARNT-RAT ASC1-HUMAN ASC1-MOUSE ASC1-RAT ASC1-XENLA ASC2-HUMAN ASC2-MOUSE ASC2-RAT AST3-DROME AST4-DROME AST5-DROME AST8-DROME ATH1-HUMAN ATH1-MOUSE NDF6-MOUSE NDF4-MOUSE NDF4-XENLA NGN2-MOUSE NGN3-MOUSE ATO-DROME BET3-MESAU BMAL-HUMAN CBF1-KLULA CLOC-DROME CLOC-HUMAN CLOC-MOUSE CYCL-DROME DEI-DROME DPN-DROME EMC-DROME ESC1-SCHPO ESM3-DROME ESM5-DROME ESM8-DROHY ESM8-DROME ESM8-DROME ESMC-DROME ESMC-DROME ESMC-DROME HAIR-DROME HAIR-DROVI HANI-CHICK HANI-HUMAN HANI-MOUSE HANI-RAT HANI-SHEEP HANI-XENLA HAN2-BRARE HAN2-CHICK HAN2-HUMAN HAN2-MOUSE HAN2-XENLA HEN1-HUMAN HEN1-MOUSE HEN2-HUMAN HES1-CHICK HES1-HUMAN HES1-MOUSE HES1-RAT HES2-HUMAN HES2-MOUSE HES2-RAT HES3-RAT HES5-MOUSE HES5-RAT HEY1-CANFA HEY1-HUMAN HEY1-MOUSE HIFA-HUMAN HIFA-MOUSE HTF4-CHICK HTF4-HUMAN HTF4-MESAU HTF4-MOUSE HTF4-PAPHA HTF4-RAT HTF4-XENLA ID1-HUMAN ID1-MOUSE ID1-RAT ID2-HUMAN ID2-MOUSE ID2-RAT ID3-HUMAN ID3-MOUSE ID3-RAT ID4-HUMAN ID4-MOUSE INO2-YEAST INO4-YEAST ITF2-CHICK ITF2-CHICK ITF2-HUMAN ITF2-MOUSE ITF2-RAT LYL1-HUMAN LYL1-MOUSE MAD4-HUMAN MAD-HUMAN MAD-MOUSE MAX-BRARE MAX-CHICK MAX-HUMAN MAX-MOUSE MAX-RAT MAX-XENLA MF25-XENLA MITF-HUMAN MITF-RAT MLX-HUMAN MLX-MOUSE MNT-HUMAN MNT-MOUSE MUSC-HUMAN MUSC-MOUSE MX11-HUMAN MX11-MOUSE MX11-RAT MYC1-CYPCA MYC1-XENLA MYC2-CYPCA MYC2-MARMO MYC2-SPEBE MYC2-XENLA MYC-B-RAT MYC-L-HUMAN MYC-L-MOUSE MYC-L-XENLA MYCM-HUMAN MYCM-XENLA MYCN-CHICK MYCN-HUMAN MYCN-MARMO MYCN-MOUSE MYCN-RAT MYCN-SERCA MYCN-XENLA MYCS-RAT MYC-ASTVU MYC-AVIM2 MYC-AVIMC MYC-AVIMD MYC-AVIME MYC-AVIOK MYC-BRARE MYC-CALJA MYC-CANFA MYC-CARAU MYC-CHICK MYC-FELCA MYC-FLVTT MYC-HUMAN MYC-MARMO MYC-MOUSE MYC-ONCMY MYC-PANTR MYC-PIG MYC-RAT MYC-SHEEP MYF5-BOVIN MYF5-CHICK MYF5-COTJA MYF5-HUMAN MYF5-MOUSE MYF5-NOTVI MYF5-XENLA MYF6-CHICK MYF6-MOUSE MYF6-RAT MYF6-XENLA MYO1-ONCMY MYO2-ONCMY MYO2-CAEER MYO2-CAEEL MYO2-CHICK MYO2-COTJA MYO2-DROME MYO2-MOUSE MYO2-PIG MYO2-RAT MYO2-SHEEP MYO2-XENLA MYOG-CHICK MYOG-COTJA MYOG-HUMAN MYOG-MOUSE MYOG-PIG MYOG-RAT NDF1-CHICK NDF1-HUMAN NDF1-MOUSE NDF1-RAT NDF1-XENLA NDF2-HUMAN NDF2-MOUSE NDF2-RAT NGN1-HUMAN NGN1-MOUSE NGN1-RAT NDFM-CHICK NPA1-HUMAN NPA1-MOUSE NPA2-HUMAN NPA2-MOUSE NUC1-NEUCR PAS1-HUMAN PAS1-MOUSE PHO4-YEAST OLG2-HUMAN RTG1-YEAST RTG3-YEAST SCL-CHICK SIM1-HUMAN SIM2-HUMAN SIM2-MOUSE SIMA-DROME SIM-DROME SIM-DROME TWST-DROME TWST-MOUSE TWST-XENLA TYE7-YEAST USF1-HUMAN USF1-MOUSE USF1-RABIT USF1-XENBO USF2-HUMAN USF2-MOUSE USF-STRPU WS14-HUMAN WS14-MOUSE YAWC-SCHPO YLB7-CAEEL YMH7-CAEEL YNP2-CAEEL YRY3-CAEEL TWST-CAEEL</b></p>
PS00061	<p><b>2BHD-STREX ADH2-DROMN ADH-DROMA ADH-DROMM ADH-DROSL DECR-RAT DHB7-RAT DHGA-BACME DHG-BACSU DH2-RABIT DH2-SHEEP DHK2-STRVN ENTA-ECOLI MASI-AGRT9 PGDH-HUMAN Y019-THEMA YAEB-SCHPO YF43-MYCTU YWC4-CAEEL OXIR-STRAT 25KD-SARPE 3BHD-COMTE ACT3-STRCO ADH1-CERCA ADH1-DROHY ADH1-DROMN ADH1-DROMO ADH1-DROMT ADH1-DROMU ADH1-DRONA ADH2-CERCA ADH2-DROAR ADH2-DROBU ADH2-DROHY ADH2-DROMO ADH2-DROMU ADH-DROMY ADH2-DROWH ADHR-DROAM ADHR-DROER ADHR-DROGU ADHR-DROIM ADHR-DROLE ADHR-DROMA ADHR-DROMD ADHR-DROME ADHR-DROPE ADHR-DROPS ADHR-DROSU ADHR-DROAM ADHR-DROYA ADH-BACOL ADH-DROAD ADH-DROAF ADH-DROAM ADH-DROBE ADH-DROCR ADH-DRODI ADH-DROER ADH-DROFL ADH-DROGR ADH-DROGU ADH-DROHA ADH-DROHE ADH-DROIM ADH-DROLA ADH-DROLE ADH-DROMD ADH-DROME ADH-DRONI ADH-DROOR ADH-DROPE ADH-DROPI ADH-DROPL ADH-DROPS ADH-DROSI ADH-DROSU ADH-DROTE ADH-DROTS ADH-DROWI ADH-DROYA ADH-SCAAL ADH-ZAPTU ARDH-CANAL ARDH-CANTR ARDH-PICTST BA71-EUBSP BA72-EUBSP BDHA-ALCEU BDHA-RHIME BDH-BOVIN BDH-HUMAN BDH-RAT BEND-ACICA BLI4-NEUCR TODD-PSEPU BPHB-BURCE BPHB-COMTE BPHB-PSEPS BPHB-PSEPU BPHB-PSES1 BPHB-RHOGO BUDC-KLEPN BUDC-KLETE CBR2-CAEEL CBR2-MOUSE CBR2-PIG CMTB-PSEPU DECR-HUMAN DHB1-HUMAN DHB1-MOUSE DHB1-RAT DHB2-HUMAN DHB2-MOUSE DHB2-RAT DHB3-HUMAN DHB3-MOUSE DHB3-RAT DHB4-HUMAN DHB4-MOUSE DHB4-RAT DHB7-HUMAN DHB7-MOUSE DHB8-CALJA DHB8-HUMAN DHB8-MOUSE DHB8-MOUSE DHB8-MOUSE DHBV-CAEEL DHBV-CAEEL DHBX-ANAPL DHBX-CAEEL DHBX-CAEEL DHC3-HUMAN DHCA-HUMAN DHCA-MOUSE DHCA-RABIT DHCA-RAT DHG1-BACME DHG2-BACME DHG2-BACSU DHG3-BACME DHG4-BACME DHGB-BACME DHG-BACME DH1-HUMAN DH1-MOUSE DH1-RAT DH1-SAIS DH1-SHEEP DH2-BOVIN DH2-HUMAN DH2-MOUSE DH2-RAT DHK1-STRVN DHKR-STRCM DHMA-FLA51 DHPH-HUMAN DHPH-RAT DHSO-RHOSH DIDH-COMTE DIDH-PSEPD DLTE-BACSU EPHD-MYCTU FABG-ACTAC FABG-AQUAE FABG-ARATH FABG-BACSU FABG-BRANA FABG-BUCAI FABG-CHLMA FABG-CHLPM FABG-CHLTR FABG-CUPLA FABG-ECOLI FABG-HAEN FABG-MYCAV FABG-MYCSM FABG-MYCTU FABG-PERAE FABG-PSEAE FABG-RICPR FABG-SALTY FABG-VIBCH FABG-VIBHA FAG1-SYNY3 FAG2-SYNY3 FBP2-DROME FIXR-BRAJA FOX2-CANTR FOX2-NEUCR FOX2-YEAST FVT1-HUMAN GNO-GLUOX GS39-BACSU HCAB-ECOLI HCD2-BOVIN HCD2-DROME HCD2-HUMAN HCD2-MOUSE HCD2-RAT HDHA-CLOSO HDHA-ECOLI DHS2-HUMAN HETN-ANASP IDNO-ECOLI KDUD-BACSU KDUD-ECOLI KDUD-ERWCH L767-CAEEL LGID-PSEPA LINC-PSEPA LINX-PSEPA MOAE-KLEAE MS11-AGRRH MS12-AGRRH MTDH-UROFA NAHB-PSEPU NODG-AZOBR NODG-RHIME NODG-RHIS3 NOG4-RHIME NORI-ASPPA OXIR-STRLI PHAB-ACISP PHAB-PARDE PHBB-ALCEU PHBB-CHRV1 PHBB-RHIME PHBB-ZOORA PTMA-CAMCO PTR1-LEIMA PTR1-LEITA RDH1-BOVIN RDH1-HUMAN RHLG-PSEAE RIDH-KLEAE ROH1-RAT ROH2-RAT ROH3-RAT SDR1-PICAB SORD-KLEPN SOU1-CANAL SOU2-CANAL SP19-YEAST SRLD-ECOLI STCE-EMENI STCU-EMENI T4HR-MAGGR TODD-PSEPU TRN1-DATST TRN2-DATST TRN2-HYONI TRNH-DATST TS2-MAIZE UCPA-ECOLI UCPA-SALTY VDLCL-HELPJ VDLCL-HELPY VER1-ASPPA VIBA-VIBCH XYLL-PSEPU Y048-HAEN Y1J1-MYCTU Y325-THEMA Y432-LISMO Y452-LISIN Y484-MYCTU Y4EK-RHISN Y4EL-HYCN Y4LA-RHISN Y4PM-RHISN Y4VI-RHISN Y945-MYCTU YAY8-SCHPO YB09-YEAST YB45-SCHPO YBBO-ECOLI YCIK-ECOLI YL46-BRAJA YD1F-SCHPO YD50-MYCTU YDFG-BACNO YDFG-ECOLI YDFG-HAEN YDFG-SALTY YDGB-ECOLI YGCV-ECOLI YGFF-ECOLI YGHA-ECOLI YHDF-BACSU YHXC-BACSU YHXC-BACSU YIM4-YEAST YIV5-YEAST YIV6-YEAST YJGI-ECOLI YK02-MYCTU YK73-MYCTU YKF5-YEAST YKUF-BACSU YM71-YEAST YMEC-METEX YMP3-STRCO YOHF-ECOLI YOXD-BACSU YQJQ-BACSU YURA-MYXXA YUSZ-BACSU YUXG-BACSU YV06-PSEAE YVX3-CAEEL YWFD-BACSU YWFH-BACSU YXBG-BACSU YXEK-CAEEL YXJF-BACSU</b></p>

(continued)



TABLE 4. (Continued)

Family	Protein ID's
PS00301	CYSN-RHTR CYSN-XYLEA EF1A-ARCFU EF1A-DICDI EF1A-SULSO EF1S-PORPU EF2-CHICK EF2-MESAU EFTU-CHLTR EFTU-FERIS EFTU-GRALE EFTU-MYCPN EFTU-NEPOL EFTU-TOBAC EFTU-XYLEA LEPA-MYCHY LEPA-MYCLE LEPA-MYCPN TETQ-PREIN TYPA-SYNY3 CYSN-BUCAI CYSN-ECOLI CYSN-MYCTU CYSN-PSEAE CYSN-RHIME EF10-XENLA EF11-CRIGR EF11-DAUCA EF11-DROME EF11-EUPCR EF11-HORVU EF11-HUMAN EF11-MOUSE EF11-RHIRA EF11-SCHPO EF11-XENLA EF12-DAUCA EF12-DROME EF12-ABSOL EF12-EUPCR EF12-HORVU EF12-HUMAN EF12-MOUSE EF12-RHIRA EF12-SCHPO EF12-XENLA EF13-RHIRA EF12-SCHPO EF13-XENLA EF14-ABSO EF14-CAEEL EF14-AERPE EF14-AJECA EF14-APIME EF14-ARATH EF14-ARTSA EF14-ARXAD EF14-ASHGO EF14-AURPU EF14-BLAHO EF14-BOMMO EF14-BRARE EF14-CAEEL EF14-CANAL EF14-CHICK EF14-CRYNE EF14-CRYPV EF14-DESMO EF14-EMBO EF14-ENTHI EF14-EUCGR EF14-GIALA EF14-HALHA EF14-HALMA EF14-HELVI EF14-HYDAT EF14-LYCES EF14-MAIZE EF14-MANES EF14-METJA EF14-METTH EF14-METWA EF14-NEUCR EF14-ONCVO EF14-ORYSA EF14-PEA EF14-PLAFK EF14-PODAN EF14-PODCU EF14-PUOCR EF14-PYRAB EF14-PYRAE EF14-PYRHO EF14-PYRWO EF14-RHYAM EF14-SCHCO EF14-SORMSA EF14-SOYBN EF14-STYLE EF14-SULAC EF14-TETPY EF14-THEAC EF14-THECE EF14-TOBAC EF14-TRIRE EF14-TRYBB EF14-VICFA EF14-WHEAT EF14-YARLI EF14-YEAST EF1C-PORPU EF2-AERPE EF2-ARCFU EF2-BETVU EF2-BLAHO EF2-CAEEL EF2-CANAL EF2-CHLKE EF2-CRIGR EF2-CRYPV EF2-DESMO EF2-DICDI EF2-DROME EF2-ENTHI EF2-HALHA EF2-HUMAN EF2-METBU EF2-METJA EF2-METMT EF2-METTE EF2-METTH EF2-METVA EF2-MOUSE EF2-PYRAB EF2-PYRHO EF2-PYRFU EF2-RABIT EF2-RAT EF2-SCHPO EF2-SULAC EF2-SULSO EF2-THEAC EF2-THECE EF2-TYMT EFG1-BORBU EFG1-STRCO EFG1-SYNY3 EFG1-TREPA EFG1-YEAST EFG2-BORBU EFG2-STRCO EFG2-SYNY3 EFG2-TREPA EFG2-YEAST EFGC-PEA EFGC-SOYBN EFGM-MYCTU EFGM-SYNY3 EFGM-TREMA EFG1-RAT EFG-AGRTU EFG-APPFP EFG-AQUAE EFG-AQUPY EFG-BACHD EFG-BACST EFG-BACSU EFG-BUCAI EFG-CHLMU EFG-CHLPH EFG-CHLTR EFG-ECOLI EFG-HAEN EFG-HELPI EFG-HELPU EFG-MICLU EFG-MYCGE EFG-MYCLE EFG-MYCPN EFG-MYCTU EFG-NEIGO EFG-PASMU EFG-PLARO EFG-RICCN EFG-RICPR EFG-SALTY EFG-SPIPL EFG-STIAM EFG-STRPY EFG-STRRA EFG-SYNP6 EFG-THEMA EFG-THETH EFG-THICU EFG-UREPA EFT1-SOYBN EFT1-STRCO EFT1-STRCU EFT1-STRRA EFT2-SOYBN EFT2-STRRA EFT3-STRCO EFT3-STRRA EFT4-PASMU EFT4-PASMU EFTU-AGRTU EFTU-APPFP EFTU-AQUAE EFTU-AQUIP EFTU-ARATH EFTU-ASTLO EFTU-BACFR EFTU-BACHD EFTU-BACST EFTU-BACSU EFTU-BORBU EFTU-BOVIN EFTU-BRELN EFTU-BRYPL EFTU-BUCAI EFTU-BUCAP EFTU-BUCMH EFTU-BUCSC EFTU-BURCE EFTU-CAMJE EFTU-CHACO EFTU-CHLAU EFTU-CHLMU EFTU-CHLRE EFTU-CHLPI EFTU-CHLVI EFTU-CHLVU EFTU-CODFR EFTU-COLOB EFTU-CORGL EFTU-COSCS EFTU-CYAPA EFTU-CYME EFTU-CYTLU EFTU-DEIRA EFTU-DEISP EFTU-DERMA EFTU-ECOLI EFTU-EIKKO EFTU-EUGGR EFTU-FIBUS EFTU-FLAFE EFTU-FLESI EFTU-GLOS1 EFTU-GLOVI EFTU-GONPE EFTU-GUITH EFTU-GUITH EFTU-HAEN EFTU-HELPI EFTU-HELPU EFTU-HERAU EFTU-HUMAN EFTU-MANSQ EFTU-MESVI EFTU-MICLU EFTU-MYCGA EFTU-MYCGE EFTU-MYCHO EFTU-MYCLE EFTU-MYCTU EFTU-NEIGO EFTU-ODOSI EFTU-PANMO EFTU-PEA EFTU-PHOEC EFTU-PLARO EFTU-PLEBO EFTU-PORPU EFTU-PROHO EFTU-PSEAE EFTU-RECAM EFTU-RHILU EFTU-RICPR EFTU-SALTY EFTU-SALTY EFTU-SHEPU EFTU-SPIAU EFTU-SPIPL EFTU-STIAU EFTU-STRAU EFTU-STRCJ EFTU-STRLU EFTU-STRMU EFTU-STROK EFTU-STRPY EFTU-SYNP6 EFTU-SYNP7 EFTU-SYNY3 EFTU-TAXOC EFTU-THEAQ EFTU-THEMA EFTU-THETH EFTU-THICU EFTU-TREHY EFTU-TREPA EFTU-UREPA EFTU-WOLSU EFTU-YEAST EFTU-YECPAN EFT2-PCIP1 ERK2-SCHPO ERK2-YEAST GSP1-HUMAN GUF1-YEAST HBS1-YEAST LEPA-AQUAE LEPA-BACHD LEPA-BACSU LEPA-BORBU LEPA-BORPE LEPA-BUCAI LEPA-CHLMU LEPA-CHLPI LEPA-CHLTR LEPA-ECOLI LEPA-HAEN LEPA-HELPI LEPA-HELPU LEPA-LACLA LEPA-MYCGE LEPA-MYCTU LEPA-PSEEL LEPA-RICPR LEPA-SALTY LEPA-STRCO LEPA-SYNY3 LEPA-THEMA LEPA-TREPA NODQ-AZOBK NODQ-RHIME NODQ-RHIS3 NODQ-RHISB NODQ-RHINR OTRA-STRRM RFB-BACNO RFB3-BUCAI RFB3-ECOLI RFB3-HAEN RFB3-LACLA RFB3-PASMU RFB3-SALTY RFB3-STAU RFB3-SYNY3 RFB3-DESBA SELB-ECOLI SELB-HUMAN SELB-METJA SELB-MOOTH SELB-MOUSE SN14-YEAST TET1-ENTFA TETS-ENTFA TET9-ENTFA TETM-NEIME TETM-STAAU TETM-STRLI TETM-STRPN TETM-UREUR TETO-CAMCO TETO-CAMJE TETO-STRMU TETO-STRPN TETP-CLOPE TETQ-BACFR TETQ-BACIN TETQ-CACTN TETQ-PRERU TETS-LACLA TETS-LISMO TETW-BUTHI TYPA-BACSU TYPA-BUCAI TYPA-ECOLI TYPA-HAEN TYPA-HELPI TYPA-HELPU USS1-HUMAN USS1-MOUSE YE14-SCHPO YNQ3-YEAST Y081-CAEEL

TABLE 5. DESCRIPTION OF THE PROSITE 2 DATASET

Family	Protein ID's
PS00070	DHAE-MACPR DHAX-HUMAN DHA1-BOVIN HPCCE-ECOLI YH9J-YEAST GABD-ECOLI MAOC-ECOLI DHA4-YEAST DHA3-BACSU DHA5-YEAST YLQ6-CAEEL DHAS-CHICK DHAM-BOVIN PUT2-HUMAN MMSA-CAEEL ALDA-ECOLI ALDB-ECOLI ASTD-ECOLI ASTD-PSEAE CALB-CAUCR CALB-PSEAE CALB-PSESP CROM-OCTDO CROM-OMMSL DHA1-BACSU DHA1-CHICK DHA1-ENTHI DHA1-HORSE DHA1-HUMAN DHA1-MOUSE DHA1-RAT DHA1-SHEEP DHA2-ALCEU DHA2-BACST DHA2-BACSU DHA2-HUMAN DHA2-MOUSE DHA2-RAT DHA2-YEAST DHA3-YEAST DHA4-HUMAN DHA4-MOUSE DHA4-RAT DHA5-BOVIN DHA5-HUMAN DHA6-HUMAN DHA6-YEAST DHA7-HUMAN DHA8-HUMAN DHA9-POLMI DHAB-AMAHF DHAB-ATRHO DHAB-BACSU DHAB-BETVU DHAB-ECOLI DHAB-GADCA DHAB-HORVU DHAB-ORYSA DHAB-RHIME DHAB-SPIOL DHAC-RAT DHAE-ELFED DHAF-VIBHA DHAG-HUMAN DHAG-PIG DHAL-AGABI DHAL-ALITAL DHAL-ASPNG DHAL-BACST DHAL-CLAHE DHAL-DEIRA DHAL-ECOLI DHAL-EMENI DHAL-ENCUB DHAL-MYCTU DHAL-PSEOL DHAL-PSESP DHAL-RHIOU DHAL-STRCO DHAL-VIBCH DHAM-HUMAN DHAM-MOUSE DHAM-LEITA DHAM-MESAU DHAM-MOUSE DHAM-RAT DHAN-MACPR DHAP-BOVIN DHAP-HUMAN DHAP-MOUSE DHAP-RAT DHAX-PEA DHAX-YEAST DHAY-YEAST DMPC-PSESP FEAB-ECOLI FTDH-HUMAN FTDH-RAT GABD-DEIRA GABD-RHISN GABD-SYNY3 GAPN-MAIZE GAPN-NICLE GAPN-PEA GAPN-STRMU MMSA-BACSU MMSA-BOVIN MMSA-HUMAN MMSA-PSEAE MMSA-RAT NAHF-PSESP PUT2-AGABI PUT2-YEAST PUTA-ECOLI PUTA-KLEAE PUTA-RHIME PUTA-SALTY ROCA-BACSU SSDH-HUMAN SSDH-RAT THCA-RHOER UGA5-YEAST XYC2-ACIGB XYLC-PSEPU XYLG-PSEPU Y4UC-RHISN YDCW-ECOLI YM00-YEAST YNEI-ECOLI
PS00077	COX1-THETH COX1-BACFI COX1-DIDMA COX1-ASCSU COX1-HORSE COX1-EPHEQ FIXN-AZOCA COX1-SYNYU COX1-CRION COX1-ALLMA AOX1-AERPE COX1-PEA COX1-RHOSH COX1-SOYBN COX1-PLABE CO13-THETH CO14-BRAJA COX1-ACACA COX1-ALBCO COX1-ALBTU COX1-AMICA COX1-ANAPL COX1-ANOGA COX1-ANOQU COX1-APLI COX1-APTAU COX1-ARATH COX1-ARTSF COX1-ASTPE COX1-BACP3 COX1-BACSU COX1-BALPH COX1-BALPH COX1-BETVU COX1-BOVIN COX1-BRAJA COX1-CAEEL COX1-CANFA COX1-CANSI COX1-CAPHI COX1-CARAU COX1-CASBE COX1-CERSI COX1-CHICK COX1-CHLRE COX1-CHOB1 COX1-CHOCR COX1-CHOFU COX1-CHOOC COX1-CHORO COX1-COTIA COX1-CROLA COX1-CYACA COX1-CYPCA COX1-DASNO COX1-DINSE COX1-DROME COX1-DRONO COX1-DROYA COX1-EMENI COX1-EQUAS COX1-FELCA COX1-GADMO COX1-GEOSD COX1-GOMVA COX1-HALGR COX1-HALHA COX1-HANWI COX1-HIPAM COX1-HUMAN COX1-KLULA COX1-LATCH COX1-LEITA COX1-LEPOC COX1-LESPS COX1-LOCM1 COX1-LUMTE COX1-MACRO COX1-MAIZE COX1-MARPO COX1-MEGAT COX1-METSE COX1-MOUSE COX1-MYCTU COX1-MYTED COX1-MYXGL COX1-NEUCR COX1-NOTPE COX1-OENBE COX1-ONCMY COX1-ORXAN COX1-ORYSA COX1-PANBU COX1-PAPHA COX1-PARLI COX1-PARTE COX1-PECCA COX1-PELSU COX1-PETMA COX1-PHOVI COX1-PHYME COX1-PHYPO COX1-PIG COX1-PISOC COX1-PLACH COX1-PLAFA COX1-PODAN COX1-POLOR COX1-POLSP COX1-POLSX COX1-POPMI COX1-PONPA COX1-PROWI COX1-RABIT COX1-RAT COX1-RHEAM COX1-RHILE COX1-RHISA COX1-RHUN COX1-RHOCA COX1-RICPR COX1-SACDO COX1-SALSA COX1-SALTR COX1-SCAPL COX1-SCHPO COX1-SCYCA COX1-SHEEP COX1-SORBI COX1-SQUAC COX1-STRCA COX1-STRCU COX1-SYNY3 COX1-TETPY COX1-TINMA COX1-TRIRU COX1-TRYBB COX1-WHEAT COX1-XENLA COX1-YEAST COXN-BRAJA CX1A-PARDE CX1B-PARDE CYOB-BUCAI CYOB-ECOLI CYOB-PSEPU FIXN-AGRT7 FIXN-BRAJA FIXN-RHIME NORB-PSEAE NORB-PSEST QOX1-ACEAC QOX1-BACSU QOX1-SULAC QOXM-SULAC
PS00118	PA21-NAJMO PA21-HORSE PA2H-BUNFA PA2E-PSEAU PA2C-CRODU PA2H-BOTJR PA2C-PSEAU PA2Z-HUMAN PA22-BUNMU PA23-NAJNG PA21-TRIGA PA21-ACAAAN PA21-BOTPI PA2X-RAT PA22-PIG OC90-CAVPO OC90-HUMAN OC90-MOUSE PA20-BUNMU PA20-NOTSC PA20-PSEAU PA21-AGKHA PA21-AGKHP PA21-AGKPI PA21-BOTAS PA21-BOTJA PA21-BOTJR PA21-BOTMO PA21-BOVIN PA21-BUNMU PA21-CANFA PA21-CAVPO PA21-ERIMA PA21-HEMHA PA21-HUMAN PA21-LATSE PA21-MATBI PA21-MOUSE PA21-NAJME PA21-NAJOX PA21-NOTSC PA21-ONYSC PA21-PIG PA21-PSEAU PA21-RAT PA21-SHEEP PA21-TRIFL PA21-VIPAA PA21-VIPAZ PA22-ACAAAN PA22-AGKHA PA22-AGKHP PA22-ASPSC PA22-BITNA PA22-BOTAS PA22-BOTMO PA22-BOTPI PA22-CERGO PA22-ERIMA PA22-HELJU PA22-LATCO PA22-MATBI PA22-NAJKA PA22-NAJME PA22-NAJMO PA22-NOTSC PA22-ONYSC PA22-TRIGA PA22-TRIST PA22-VIPAZ PA23-AGKHP PA23-BOTAS PA23-BOTPI PA23-BUNMU PA23-HELJU PA23-HUMAN PA23-LATSE PA23-NAJKA PA23-NAJME PA23-NAJMO PA23-NOTSC PA23-ONYSC PA23-PSEAU PA23-TRIGA PA24-BUNMU PA24-DABRU PA24-LATSE PA24-TRIGA PA25-HUMAN PA25-MOUSE PA25-PSEAU PA25-RAT PA25-TRIGA PA25-TRIST PA26-BUNFA PA26-TRIGA PA27-DABRU PA27-TRIGA PA29-PSEAU PA2A-BUNFA PA2A-CRODU PA2A-HUMAN PA2A-MICNI PA2A-MOUSE PA2A-PSEAU PA2A-PSEPO PA2A-PSETE PA2A-RABIT PA2A-RAT PA2A-VIPAA PA2A-VIPDA PA2B-BUNFA PA2B-CRODU PA2B-MICNI PA2B-PSEPO PA2B-PSETE PA2B-TRIFL PA2B-TRIMU PA2B-VIPAA PA2C-MOUSE PA2C-PSETE PA2C-RAT PA2C-VIPAA PA2D-HUMAN PA2D-MOUSE PA2D-PSEAU PA2D-PSETE PA2E-HUMAN PA2E-MOUSE PA2F-HUMAN PA2F-MOUSE PA2G-PSEAU PA2H-AGKPI PA2H-ATRNM PA2H-LATCO PA2H-XENLA PA2I-VIPAA PA2I-VIPAZ PA2M-AGKCL PA2M-CAVPO PA2M-CROSS PA2N-BUNFA PA2N-CROSS PA2N-ECHCA PA2N-VIPAA PA2X-BUNFA PA2X-HUMAN PA2X-MOUSE PA2X-NOTSC PA2X-TRIFL PA2Y-HUMAN PA2Y-MOUSE PA2Y-TRIFL PA2Z-MOUSE PA2A-AIPLA PA2A-APIME PA2A-BITCA PA2A-BITGA PA2A-BOMTE PA2A-CERCE PA2A-CROAD PA2A-CROAT PA2A-DABRR PA2-ENHSC PA2-HELHO PA2-LATLA PA2-NAJAT PA2-NAJNA PA2-OPHHA PA2-RHONO PA2-TRIOK PA2-VIPBB
PS00180	GLNA-COLGL GLN4-PEA GLN2-DROME GLNA-HELPU GLNA-PANAR GLN3-RHILP GLN1-ARATH GLN5-MAIZE GLNA-PIG GLNA-PYRHO GLNA-THIFE GLNA-SALTY GLN3-PHAVU GLNA-NICPL GLN2-DAUCA GLN1-ALNGL GLN1-BRAJA GLN1-CHLRE GLN1-DAUCA GLN1-DROME GLN1-FRAAL GLN1-LOTIA GLN1-MAIZE GLN1-MEDSA GLN1-MYCTU GLN1-ORYSA GLN1-PEA GLN1-PHAVU GLN1-RHILV GLN1-RHIME GLN1-SOYBN GLN1-STRPR GLN1-STRYR GLN1-VITVI GLN2-ARATH GLN2-BRAJA GLN2-CHLRE GLN2-FRAAL GLN2-HORVU GLN2-MAIZE GLN2-MEDSA GLN2-MYCTU GLN2-ORYSA GLN2-PEA GLN2-PHAVU GLN2-RHILP GLN2-RHIME GLN2-SOYBN GLN2-STRHY GLN2-STRYR GLN2-VITVI GLN3-HORVU GLN3-LUPAN GLN3-MAIZE GLN3-MEDSA GLN3-ORYSA GLN3-PEA GLN3-RHIME GLN4-MAIZE GLN4-PHAVU GLNA-AGABI GLNA-ANASP GLNA-AQUAE GLNA-ARCFU GLNA-AZOBK GLNA-AZOCA GLNA-AZOV1 GLNA-BACCE GLNA-BACFR GLNA-BACSU GLNA-BOVIN GLNA-BUTPI GLNA-CAEEL GLNA-CHICK GLNA-CLOSA GLNA-CRIOU GLNA-DUNSA GLNA-ECOLI GLNA-FREDI GLNA-HAEN GLNA-HALNI GLNA-HALVO GLNA-HELPI GLNA-HUMAN GLNA-LACDE GLNA-LACLA GLNA-LACSA GLNA-LUPLU GLNA-METCA GLNA-METJA GLNA-METMP GLNA-METTH GLNA-METVO GLNA-MOUSE GLNA-NEIGO GLNA-PASMU GLNA-PINSY GLNA-PROVI GLNA-PYRAB GLNA-PYRFU GLNA-PYRKO GLNA-PYRWO GLNA-RHIOU GLNA-RHOCA GLNA-RHOSH GLNA-SCHPO GLNA-SQUAC GLNA-STAAU GLNA-STRCO GLNA-SULAC GLNA-SULSO GLNA-SYNP2 GLNA-SYNY3 GLNA-THEMA GLNA-TRITH GLNA-VIBAL GLNA-VIBCH GLNA-VIGAC GLNA-XENLA GLNA-YEAST GLNC-BRANA GLNC-MAIZE YCJF-ECOLI

(continued)







TABLE 6. (Continued)

Subfamily	Protein ID's
Prostanoid	O00326 PD2R-MOUSE PE22-MOUSE PE23-BOVIN PE23-HUMAN PE23-RABIT PF2R-MOUSE P12R-BOVIN Q9R261 TA2R-BOVIN O00325 O15191 O35932 O46657 O75228 PD2R-HUMAN PE21-HUMAN PE21-MOUSE PE21-RAT PE22-CANFA PE22-HUMAN PE22-RAT PE23-MOUSE PE23-PIG PE23-RAT PE24-HUMAN PE24-MOUSE PE24-RABIT PE24-RAT PF2R-BOVIN PF2R-HUMAN PF2R-RAT PF2R-SHEEP P12R-HUMAN P12R-MOUSE P12R-RAT Q9BGL8 Q9D627 Q9TU16 TA2R-CERAE TA2R-HUMAN TA2R-MOUSE TA2R-RAT
Nucleotide-like	AA1R-BOVIN AA1R-RAT AA2A-RAT O57466 P2Y3-MELGA P2Y6-HUMAN P2YR-RAT Q99MT6 Q9ERK9 Q9HIC0 AA1R-CANFA AA1R-CAVPO AA1R-CHICK AA1R-HUMAN AA1R-RABIT AA2A-CANFA AA2A-CAVPO AA2A-HUMAN AA2A-MOUSE AA2B-CHICK AA2B-HUMAN AA2B-MOUSE AA2B-RAT AA3R-CANFA AA3R-HUMAN AA3R-RABIT AA3R-RAT AA3R-SHEEP GPRZ-HUMAN GPRZ-MOUSE O00398 O08766 O35811 P2UR-HUMAN P2UR-MOUSE P2UR-RAT P2Y3-CHICK P2Y4-HUMAN P2Y5-CHICK P2Y5-HUMAN P2Y6-RAT P2Y8-XENLA P2Y9-HUMAN P2YR-BOVIN P2YR-CHICK P2YR-HUMAN P2YR-MELGA P2YR-MOUSE Q9BXA5 Q9BXC1 Q9BYU4 Q9CFZ4 Q9DE05 Q9JIS7 Q9N1U0 Q9PU18 Q9R202 Q9W6C4

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### **Author**

**Right running head okay as shown (short title)?**

### **Pub/Author**

**Appendix tables (Tables 4–6) were able to be picked up from electronic file for manipulation. So, these tables are supplied as type rather than art. Okay?**